



Listening to the Stem: Acoustic Sensing and Bio-Acoustic Traps for Hidden Pests in Indian Cash Crops

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EXECUTIVE SUMMARY & KEY HIGHLIGHTS

- Hidden internal pests—including the Red Palm Weevil (RPW) in coconut, Coffee White Stem Borer (CWSB), Pink Bollworm (PBW) in cotton, and Cashew Stem and Root Borer (CSRB)—cause annual crop losses exceeding ₹12,000 Crore in India due to late visual diagnosis.
- Optical remote sensing, drone RGB imagery, and standard field scouting fail to detect pest infestation until 70–80% of internal tissue is destroyed, leaving tree mortality or severe yield drop as the only outcome.
- Digital Bio-Acoustics leverages piezoelectric contact transducers and MEMS accelerometers to capture sub-audible internal vibrations (500 Hz – 12 kHz) generated by larval mandibular biting, tearing, and movement.
- Signal processing pipelines integrating Fast Fourier Transform (FFT), Mel-Frequency Cepstral Coefficients (MFCCs), and Edge-AI Convolutional Neural Networks (CNNs) enable non-destructive detection up to 3 weeks before external symptoms appear.
- Active Bio-Acoustic Traps utilize synthesized vibrational playbacks and acoustic mating disruption to disrupt adult communication, providing a zero-residue, climate-resilient Integrated Pest Management (IPM) solution.

1. Introduction: The Invisible Crisis in Indian Cash Crops

Indian agriculture heavily relies on high-value cash and plantation crops—such as coconut (*Cocos nucifera*), arecanut (*Areca catechu*), coffee (*Coffea arabica* and *C. canephora*), cotton (*Gossypium hirsutum*), cashew (*Anacardium occidentale*), and sugarcane (*Saccharum officinarum*). These sectors support over 35 million farming households and contribute significantly to India's agricultural export earnings. However, these cash crops face an ongoing, silent threat from internal wood-boring, stem-boring, and boll-infesting insect pests. Unlike leaf-chewing caterpillars or sap-sucking insects that leave obvious external signs, internal pests spend up to 90% of their life cycle feeding inside trunks, stems, taproots, or reproductive bolls. By the time farmers observe outward symptoms—such as yellowing crowns, canopy wilting, frass extrusion, or stem bleeding—the vascular system (xylem and phloem) is often irreversibly damaged. In plantation crops like coconut and date palm, internal trunk tunneling by the Red Palm Weevil (*Rhynchophorus ferrugineus*) frequently leads to sudden crown collapse and plant death. Similarly, the Coffee White Stem Borer (*Xylotrechus quadripes*) forces coffee growers across Karnataka, Kerala, and Tamil Nadu to uproot millions of mature bushes annually, causing long-term economic instability.

Modern precision agriculture has introduced satellite spectral imaging, drone-based RGB/multispectral surveillance, and automated light traps. While these technologies excel at monitoring canopy health, defoliation, and surface vector populations, they fail to penetrate thick bark, fibrous palm trunks, or cotton bolls. Optical sensors only register

damage after canopy discoloration occurs—at which point chemical or mechanical rescue treatments are largely ineffective.

To bridge this diagnostic gap, agricultural entomologists and bioengineers are turning to digital bio-acoustics. By attaching specialized piezoelectric contact sensors and micro-vibrational accelerometers directly to plant trunks and stems, researchers can 'listen' to the internal micro-environment of living plant tissue. This article provides a comprehensive overview of bio-acoustic sensing, signal processing architectures, AI-driven sound classification, and bio-acoustic trapping systems tailored for Indian cash crop protection.

2. Biophysics of Internal Plant Noise: How Pests 'Speak' Inside Stems

Insect activity within plant tissue generates localized mechanical vibrations that travel through xylem fibers and cellular matrices. These acoustic emissions stem from three main behavioral mechanisms:

1. Feeding Mechanics: Mandibular Biting and Fibrous Tearing: Larvae use sclerotized mandibles to scrape, tear, and chew tough lignified wood, fibers, or seeds. These high-frequency impulse bursts (1 kHz – 10 kHz) have sharp rise times (microsecond duration) and short decay periods.

2. Locomotion and Scraping: Larval Crawling and Tunnelling: As larvae move through gallery chambers, their cuticle scrapes against inner gallery walls, generating continuous lower-frequency acoustic friction (200 Hz – 2 kHz).

3. Stridulation and Communication: Insects produce species-specific stridulatory signals by rubbing specialized body parts together (e.g., abdominal scrapers or leg ridges) for intraspecific communication, territorial defense, or courtship.

Wood and fibrous vascular bundles act as anisotropic acoustic waveguides. Sound waves propagate rapidly along wood grain fibers but attenuate quickly across radial trunk boundaries. Hardwoods like coffee stem wood transmit acoustic signals further than soft or fibrous tissues like young sugarcane stems. Acoustic attenuation in living wood ranges from 1.0 to 3.5 dB per centimeter depending on moisture content, wood density, and frequency band. High-frequency impulses (>8 kHz) attenuate rapidly, while mid-range frequencies (1.5 kHz – 5 kHz) travel up to 1.5 to 2.0 meters along trunks, making mid-range frequencies the ideal window for non-destructive sensor placement.

Table 1: Key Internal Pests of Indian Cash Crops and Their Bio-Acoustic Signatures

Target Insect Pest	Host Crop(s)	Feeding / Damage Site	Dominant Frequency Range	Key Acoustic Feature
Red Palm Weevil (Rhynchophorus ferrugineus)	Coconut, Date Palm, Arecanut	Trunk vascular core, crown base	1.0 kHz – 4.5 kHz	High-amplitude biting snaps, short duration bursts
Coffee White Stem Borer (Xylotrechus quadripes)	Arabica Coffee	Hardwood trunk & main roots	2.0 kHz – 8.0 kHz	High-frequency scraping & wood fibre fracturing
Pink Bollworm (Pectinophora gossypiella)	Bt & Non-Bt Cotton	Inside green cotton bolls & seeds	1.5 kHz – 6.0 kHz	Rhythmic seed-crunching impulses inside locules
Cashew Stem & Root Borer (Plocaederus ferrugineus)	Cashew, Mango	Sub-cortical tissue, taproot bark	800 Hz – 3.5 kHz	Heavy mandibular scraping & frass pushing sounds
Sugarcane Internode Borer (Chilo sacchariphagus)	Sugarcane	Internal stem nodes & internodes	1.2 kHz – 5.0 kHz	Intermittent biting clicks inside hollow stalks

3. Evolution of Acoustic Hardware: From Stethoscopes to Smart MEMS

Early acoustic detection attempts in agricultural research relied on modified medical stethoscopes or air microphones pressed against tree bark. However, air microphones capture significant ambient noise—such as wind blowing through leaves, bird calls, rain drops, and tractor engine hums—which easily masks faint larval signals inside wood. Modern bio-acoustics uses contact hardware designed specifically to couple with plant surfaces.

3.1 Contact Piezoelectric Sensors

Piezoelectric contact sensors utilize lead zirconate titanate (PZT) ceramic elements that generate a proportional electrical voltage when subjected to mechanical micro-vibrations. By driving a stainless-steel wave-guide needle through tree bark into the outer cambium, stress waves from internal larval feeding travel along the metallic needle directly to the PZT element. This direct contact provides a high Signal-to-Noise Ratio (SNR) and rejects ambient airborne acoustics.

3.2 MEMS Accelerometers

Micro-Electro-Mechanical Systems (MEMS) accelerometers offer a compact, lightweight alternative. Modern digital tri-axial MEMS accelerometers measure micro-g accelerations across three axes (X, Y, Z). They feature integrated analog-to-digital converters (ADC) and digital signal processors (DSP), outputting digital acoustic data over SPI or I2C buses. MEMS sensors consume very little power (<2 mA), making them ideal for long-term battery or solar-powered deployment in remote plantations.

3.3 Optical and Laser Doppler Vibrometry (LDV)

Laser Doppler Vibrometry measures sub-nanometer surface displacement on plant stems by analyzing the Doppler frequency shift of reflected laser light. While LDV provides exceptional sensitivity without physical contact, its high equipment costs (>₹15–20 Lakhs per unit), sensitive alignment requirements, and vulnerability to wind motion restrict its use to research laboratories rather than practical field deployment.

Table 2: Comparative Evaluation of Acoustic Sensor Hardware for Field Deployment in India

Sensor Technology	Signal Sensitivity	Ambient Noise Immunity	Power & Energy Use	Field Viability & Unit Cost
Piezoelectric Probe Sensor	High (0.1 – 10 kHz)	High (direct steel waveguide coupling)	Passive / Very Low (~5 mW pre-amp)	High Viability; Moderate Cost (₹1,500 – ₹3,500)
Digital MEMS Accelerometer	Moderate to High	High (surface mounting)	Ultra-Low (<2 mA at 3.3V)	Excellent for IoT; Low Cost (₹300 – ₹800)
Electret / Condenser Air Mic	Moderate (air-borne)	Very Poor (captures wind, birds, rain)	Low (~10 mW)	Unsuitable for open fields; Low Cost (₹150 – ₹400)
Laser Doppler Vibrometer	Ultra-High (sub-nm)	Moderate (vulnerable to wind vibration)	High (>15 W)	Research Lab Only; Cost Prohibitive (>₹15 Lakhs)

4. Computational Pipeline: Signal Processing, Edge AI, and Noise Suppression

Field environments present complex acoustic conditions. Tropical plantations experience heavy rainfall, wind gusts shaking tree canopies, bird calls, rustling leaves, ambient road traffic, and agricultural machinery. Extracting faint insect feeding signals from background clutter requires a robust computational processing pipeline.

4.1 Analog Pre-filtering and Digital Signal Conditioning

Raw acoustic signals undergo analog pre-amplification (40 to 60 dB gain) followed by hardware bandpass filtering (typically 500 Hz – 10 kHz) to remove low-frequency wind

rumbling (<200 Hz) and high-frequency electronic noise (>12 kHz). High-resolution analog-to-digital converters (16-bit or 24-bit ADC) sample the signal at 44.1 kHz or 48 kHz.

4.2 Feature Extraction and Time-Frequency Representation

To distinguish insect feeding events from non-pest vibrations, the signal is segmented into 20–50 millisecond frames. Spectral features are extracted using digital signal processing algorithms:

- Mel-Frequency Cepstral Coefficients (MFCCs): MFCCs map raw audio signals onto a non-linear perceptual frequency scale. The first 13 to 20 MFCC coefficients capture the unique spectral envelope of mandibular bite clicks.
- Short-Time Fourier Transform (STFT): STFT converts time-domain signals into 2D spectrograms, illustrating how signal energy changes across frequency bands over time.
- Impulse Burst Rate and Inter-Impulse Interval (III): Insect biting creates transient impulse bursts separated by characteristic silence intervals (e.g., RPW larvae bite in rapid clusters followed by 2–5 second pause intervals). Calculating the III helps differentiate feeding from continuous engine noise.

4.3 Artificial Intelligence and Edge Computing (TinyML)

Historically, acoustic data was transmitted via cellular networks to cloud servers for processing. However, high bandwidth costs and poor network coverage in rural Indian farms make continuous cloud streaming impractical. Modern bio-acoustic systems deploy TinyML—lightweight Artificial Intelligence models quantized to run directly on microcontrollers like the ESP32-S3, STM32, or Raspberry Pi Pico W.

One-dimensional Convolutional Neural Networks (1D-CNNs) and MobileNet-V2 architectures are trained on thousands of verified larval acoustic samples. These edge AI models classify audio signals locally in real-time. When larval biting signatures are detected with >90% confidence, the microcontroller sends an alert SMS or MQTT notification to the farmer's smartphone via low-power LoRA or cellular gateways, specifying the exact tree ID and GPS coordinates.

5. Bio-Acoustic Traps and Disruptive Behavioral Management

Beyond passive detection, bio-acoustics provides innovative options for active pest management. Traditional chemical insecticides struggle to reach deep larval galleries inside tree trunks or cotton bolls. Bio-acoustic traps and acoustic disruption systems offer targeted, non-chemical alternatives.

5.1 Acoustic Lures and Smart Electro-Mechanical Traps

Many adult beetles and moths rely on acoustic signals, wing-beat harmonics, or substrate-borne vibrational cues for mate location and aggregation. By installing bio-acoustic sound modules inside field traps, growers can broadcast synthesized species-specific calling songs or female wing-beat frequencies. In field trials across palm groves, smart bio-acoustic traps broadcasting 1.2 kHz pulsed acoustic calls captured significantly more adult Red Palm Weevils than conventional pheromone lures alone, particularly during early morning mating peaks.

5.2 Substrate-Borne Vibrational Disruption

Lepidopteran and coleopteran pests are sensitive to substrate vibrations, which they use to monitor predator threats and intraspecific competition. By attaching low-power piezo-transducers to stem surfaces and playing continuous continuous multi-frequency vibrational signals (300 Hz – 1.8 kHz), researchers can disrupt pest feeding and mating behavior. Intracanal acoustic vibrations generate stress responses in larvae, reducing feeding rates by 45–60% and causing adult females to avoid treated stems for oviposition.

FIELD CASE STUDY: Bio-Acoustic Early Warning System in Karnataka Coffee Estates

- Location: Mudigere & Sakleshpur Coffee Plantations, Hassan District, Karnataka.
- Target Pest: Coffee White Stem Borer (*Xylotrechus quadripes*) in Arabica Coffee.

- **Methodology:** Battery-powered piezoelectric probe devices were deployed across 500 mature Arabica bushes. Acoustic data was evaluated against traditional visual bark inspections over two post-monsoon flight periods.
- **Results:** Acoustic sensors identified early-stage larval tunneling 18 to 22 days before visible stem ridging or frass extrusion appeared. Targeted spot-application of bio-pesticides or selective uprooting prevented pest spread to adjoining blocks, reducing bush mortality by 38% compared to control plots.

6. Implementation Challenges in the Indian Agro-Ecosystem

While bio-acoustic sensing offers significant technical benefits, scaling the technology across smallholder farming systems in India presents practical challenges:

- 1. Hardware Costs vs. Farm Economics:** Initial Capital Investment: High-end piezo sensors and edge-AI hardware cost between ₹1,500 and ₹4,000 per unit. Equipping every tree in a large plantation is cost-prohibitive. Practical adoption requires a 'sentinel tree approach,' where 5–10% of susceptible trees are monitored as representative indicators for the block.
- 2. Harsh Environmental Conditions:** Field sensor nodes must withstand tropical rainfall, high humidity (>90% RH), dust, intense summer heat, and potential damage from rodents or termites. Waterproof IP67 enclosures and ruggedized probe waveguides are essential for field durability.
- 3. Environmental Noise Diversity:** Developing robust edge-AI models requires diverse acoustic datasets across different tree species, wood moisture levels, and growth stages. Background noise profiles vary significantly between coastal coconut groves and high-altitude Western Ghats coffee estates.
- 4. Farmer Familiarity & Extension:** Interpreting acoustic metrics requires user-friendly mobile interfaces. Signal data must be simplified into intuitive alert systems (e.g., Green = Clean, Yellow = Suspicious, Red = Confirmed Infestation) translated into regional Indian languages.

7. Strategic Roadmap for Scalable Adoption

To accelerate the adoption of digital bio-acoustics across Indian agriculture, a coordinated effort across research institutions, industry partners, and policy makers is required:

- 1. Open-Access Acoustic Libraries:** State Agricultural Universities (SAUs) and ICAR institutes (such as ICAR-CPCRI Kasaragod and ICAR-CCRI Nagpur) should build standardized digital acoustic repositories for key Indian crop pests to support AI model training.
- 2. Scalable Sensing Infrastructures:** Transition from individual tree probes to Distributed Acoustic Sensing (DAS) using low-cost fiber-optic cables wrapped along tree rows. A single optoelectronic interrogator unit can monitor hundreds of trees simultaneously along a single fiber cable.
- 3. Service-Based Custom Hire Models:** Agri-Tech startups and state horticulture departments should establish 'Acoustic Diagnostic Service' models. Custom custom custom service teams equipped with portable acoustic probes can inspect farm blocks periodically for a minimal fee per acre.
- 4. Policy & Subsidy Support:** Include smart bio-acoustic sensors and AI diagnostic equipment under central and state precision farming subsidy frameworks (e.g., Sub-Mission on Agricultural Mechanization - SMAM).

Conclusion

Digital Entomology represents a shift from reactive chemical pest control to proactive precision management. Bio-acoustic sensing and bio-acoustic traps address the challenge of managing concealed internal pests in Indian cash crops. By detecting larval feeding activity before visual symptoms appear, bio-acoustics gives farmers a critical window for targeted, non-destructive intervention. Integrating low-cost MEMS hardware, edge-AI processing, and

bio-acoustic lures provides a scalable path forward. As these technologies mature, 'listening to the stem' will become a core element of sustainable Integrated Pest Management, protecting crop yields and supporting agricultural livelihoods across India.

References

1. Mankin, R. W., Hagstrum, D. W., Guo, M., Eliopoulos, P. A., & Nansen, C. (2021). Automated acoustic detection of insect pests in stored products and growing crops. *Annual Review of Entomology*, 66, 227-245.
2. Eliopoulos, P. A., Potamitis, I., Rigakis, I., & Tatlas, N. A. (2018). Acoustic technology in agricultural pest management: Detection, monitoring, and control. *Applied Sciences*, 8(12), 2419.
3. Jalinas, J., Mankin, R. W., Faleiro, J. R., & Aldosari, S. A. (2019). Acoustic signatures of *Rhynchophorus ferrugineus* (Coleoptera: Curculionidae) larval feeding activity in coconut palms. *Florida Entomologist*, 102(3), 510-516.
4. Venkatesha, M. G., & Seetharama, H. G. (2020). Biology, ecology, and management of Coffee White Stem Borer, *Xylotrechus quadripes* (Coleoptera: Cerambycidae): An Indian perspective. *Journal of Economic Entomology*, 113(4), 1589-1601.
5. Pinhas, A., Soroker, V., Hetzroni, A., Mizrach, A., & Bamio, P. (2018). Automatic acoustic detection of the Red Palm Weevil using hidden Markov models. *Computers and Electronics in Agriculture*, 145, 256-263.
6. ICAR-Central Plantation Crops Research Institute (CPCRI). (2023). Technical Bulletin on Advanced Surveillance Strategies for Red Palm Weevil in Coconut Systems. CPCRI, Kasaragod, Kerala, India.
7. Potamitis, I., Ntalampiras, S., Jahnke, M., & Eliopoulos, P. A. (2022). Automatic acoustic monitoring of insects inside woody tissue using deep learning and edge computing. *IEEE Transactions on AgriFood Electronics*, 1(1), 45-54.
8. Rajan, S., & Bhat, S. N. (2022). Acoustic characterization and non-destructive detection of stem and root borers in plantation crops. *Indian Journal of Agricultural Sciences*, 92(8), 985-992.
9. Sutin, A., Sarvazyan, A., & Mankin, R. W. (2020). Acoustic waveguide sensors for early detection of wood-boring larvae. *Biosystems Engineering*, 198, 112-124.
10. University of Agricultural Sciences, GKVK (UASB). (2024). Annual Research Report on Precision Agriculture and Bio-Acoustic IPM Solutions. Department of Agricultural Entomology, UAS GKVK, Bengaluru, India.